

**Gene Discovery:**  
Approaches, Developments and Applications to Sugarcane Improvement

FAPESP BIOEN  
Workshop

18-19 March 2009



# OUTLINE

## SASRI: Genomics & Functional Genomics Research

- Landscape
  - Trajectory
- In relation to information, technological resources & research paradigms*

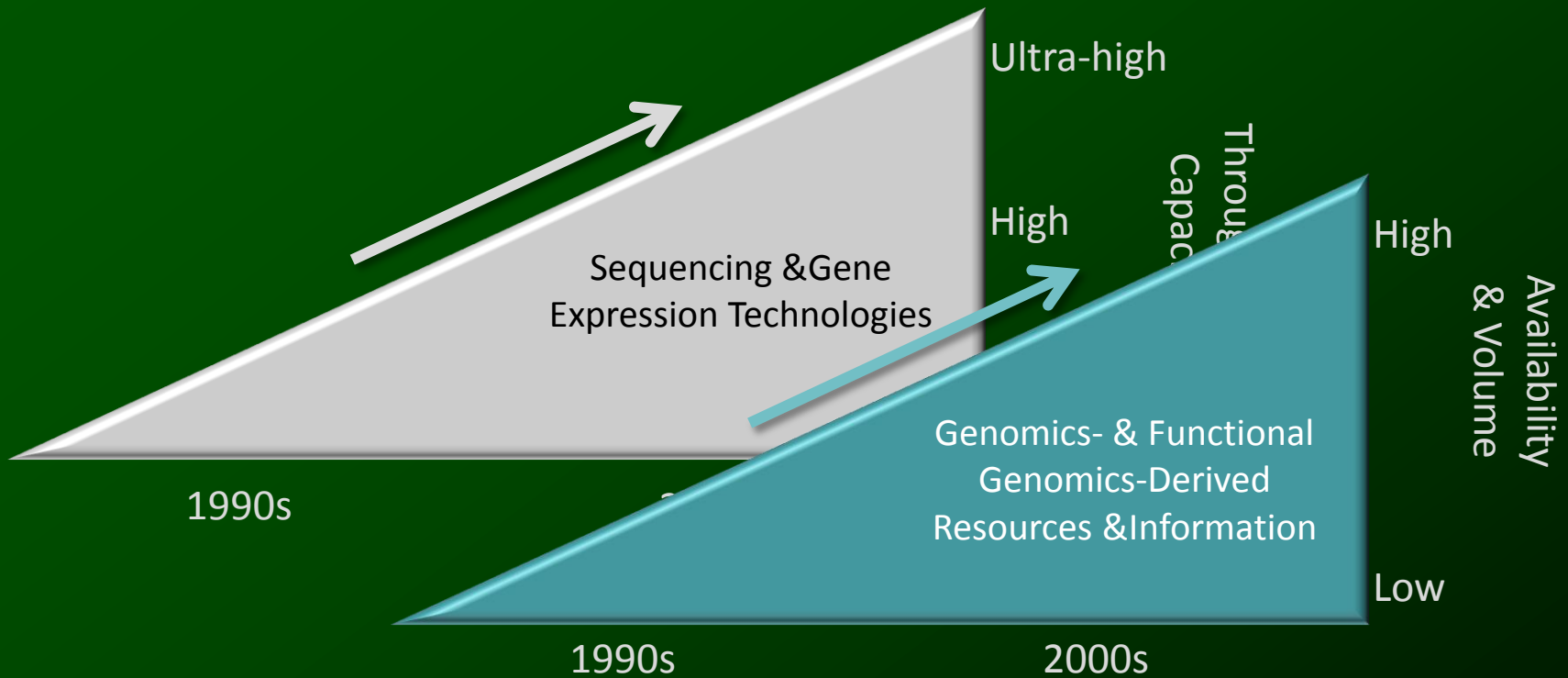
## Research: Genomics & Functional Genomics Applications

- Marker-Assisted Breeding
  - Transgenesis
  - Sugarcane Biology
- Gene sequences & their expression as access points*

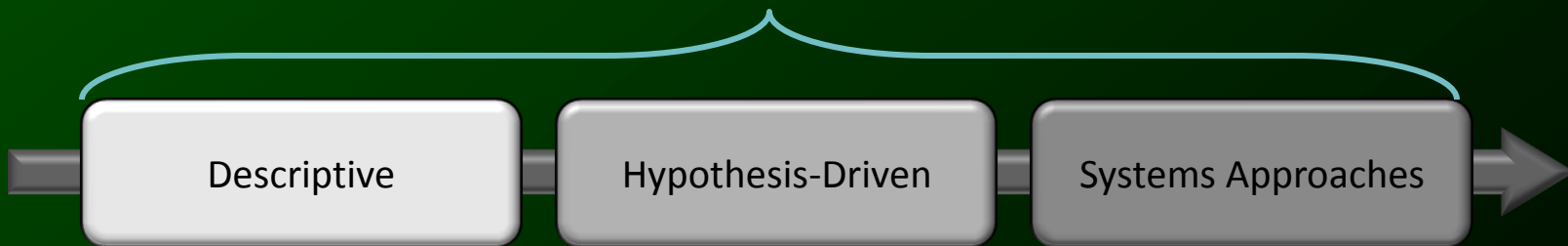
## Prospects: Genomics & Functional Genomics Advances

- Marker-Assisted Breeding
  - Sugarcane Biology
- In relation to emerging technologies & resources*

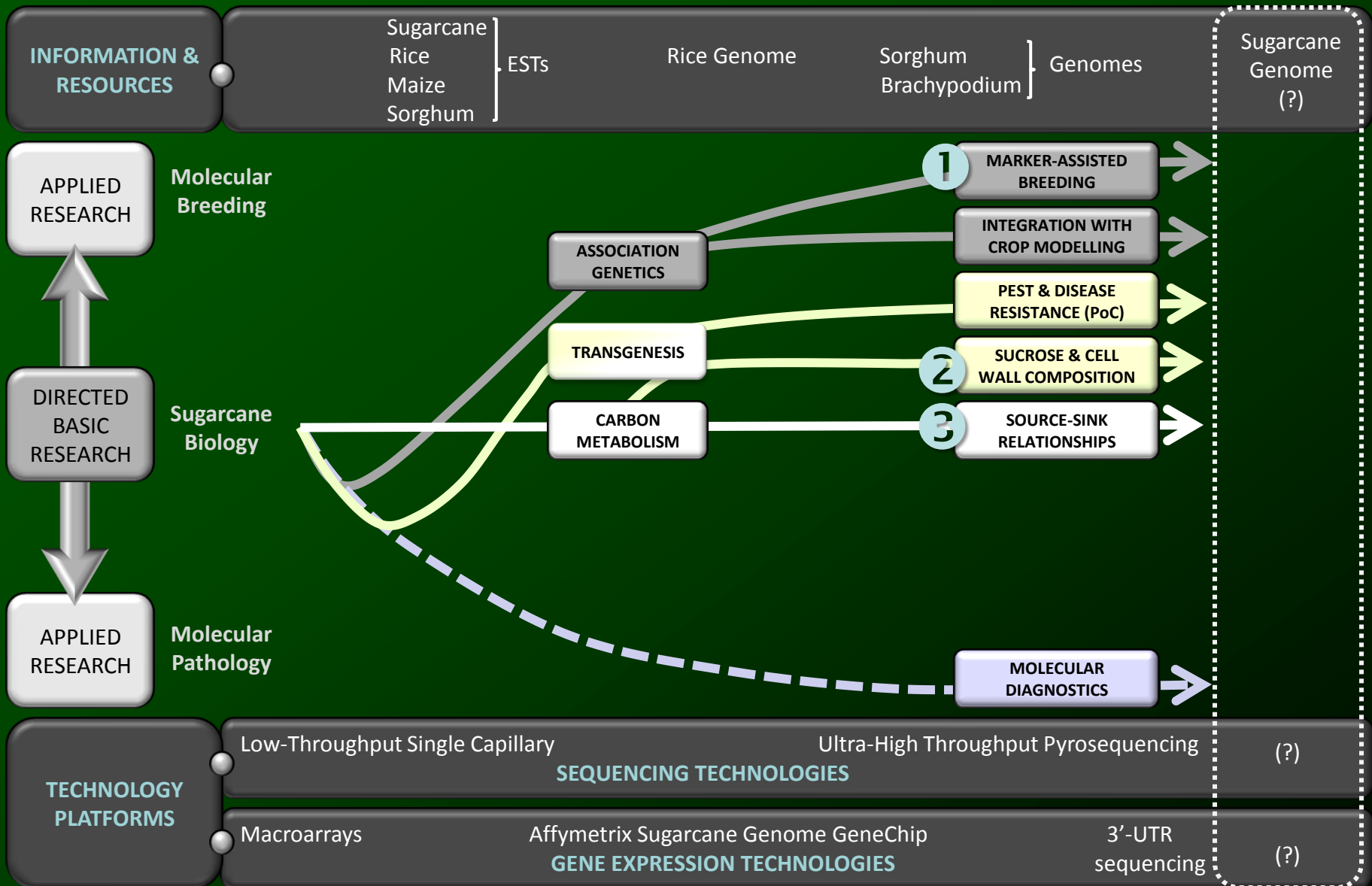
# LANDSCAPE: Resources & Research Paradigm



## RESEARCH PARADIGMS



# TRAJECTORY: From Basic to Applied Research



## CURRENT APPLICATIONS:

### TECHNOLOGIES & RESOURCES IN GENOMICS & FUNCTIONAL GENOMICS

1

**MARKER-  
ASSISTED  
BREEDING**

2

**TRANSGENESIS**

3

**SUGARCANE  
BIOLOGY  
(Source-Sink  
Relationships)**

# ASSOCIATION GENETICS: MARKER-ASSISTED BREEDING

1

## GENE SEQUENCES & THEIR EXPRESSION: MARKER DEVELOPMENT

RATIONALE & APPROACH

ISOLATION:

cDNAs differentially expressed during biotic challenge (potential resistance markers)

SSH and cDNA-AFLP differential display after pest or disease challenge

IDENTIFICATION:

sequences with putative resistance mechanism function

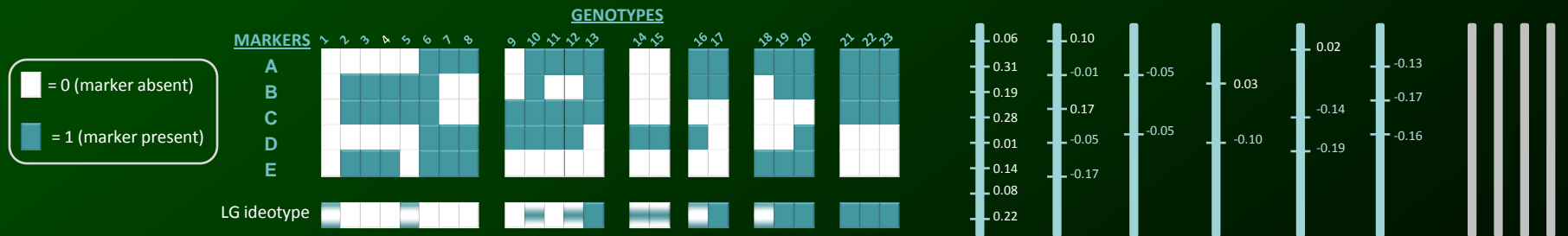
APPLICATION:

as RFLP probes on a well-characterized Linkage Disequilibrium Mapping population

numerous anonymous markers (AFLPs, SSRs) also mapped

ANALYSIS:

association mapping to identify markers of resistance/susceptibility



Haplotype Map: Example

# ASSOCIATION GENETICS: MARKER-ASSISTED BREEDING

## OUTCOMES

- Linkage Disequilibrium Map of 841 markers distributed in 231 haplotypes
- identified haplotypes/markers associated with stalk borer and smut resistance/susceptibility
- markers used to identify cross combinations predicted to give resistant progeny

SASRI: Marker-based crosses for borer and smut resistance

YEAR	NO. CROSSES
2002	79
2003	143
2004	134
2005	78
TOTAL	434

Comparison between heritability for borer resistance calculated from parent phenotype and parent marker-type (Trial of 36 bi-parental families)

	MID-PARENT PHENOTYPE	MARKER PREDICTION
$h^2$	0.560	0.820
s.e.	0.250	0.280
$r^2$	0.126	0.192
<i>P</i> -value	0.031	0.007

# TRANSGENESIS: SUCROSE & CELL WALL

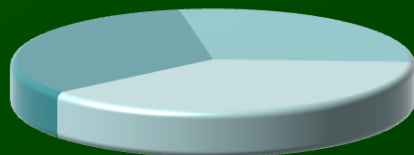
## 2 GENE SEQUENCES & THEIR EXPRESSION: ACCESSING TRANSGENES

### Sugarcane Stalk Physiology

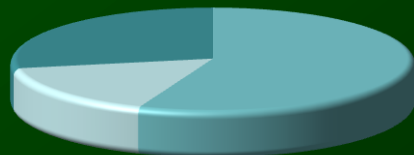
#### C-Flux



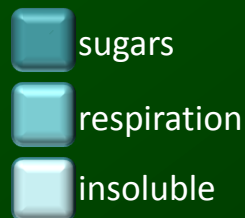
Mature internode (NCo376)  
**(HIGH SUCROSE)**



Immature internode (NCo376)  
**(LOW SUCROSE)**



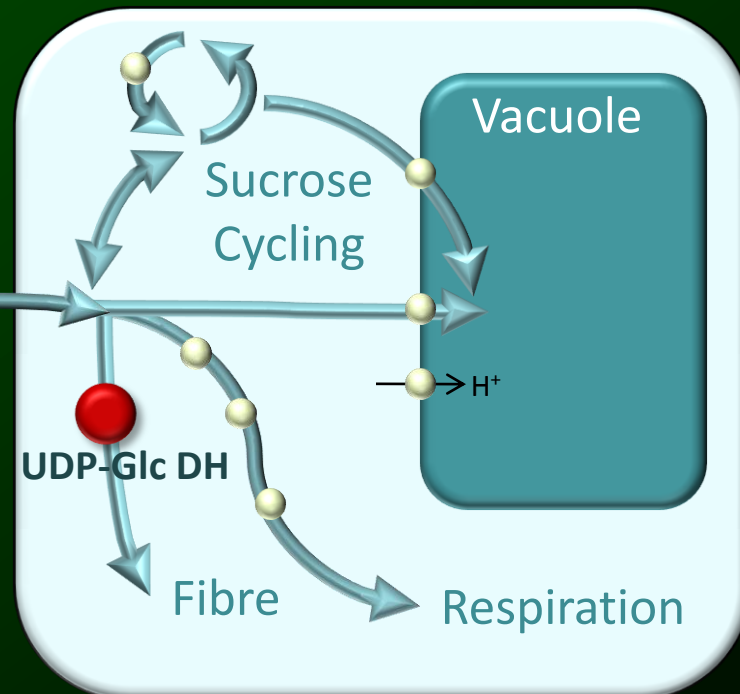
Mature internode (*S. spontaneum*)  
**(LOW SUCROSE)**



Photosynthate



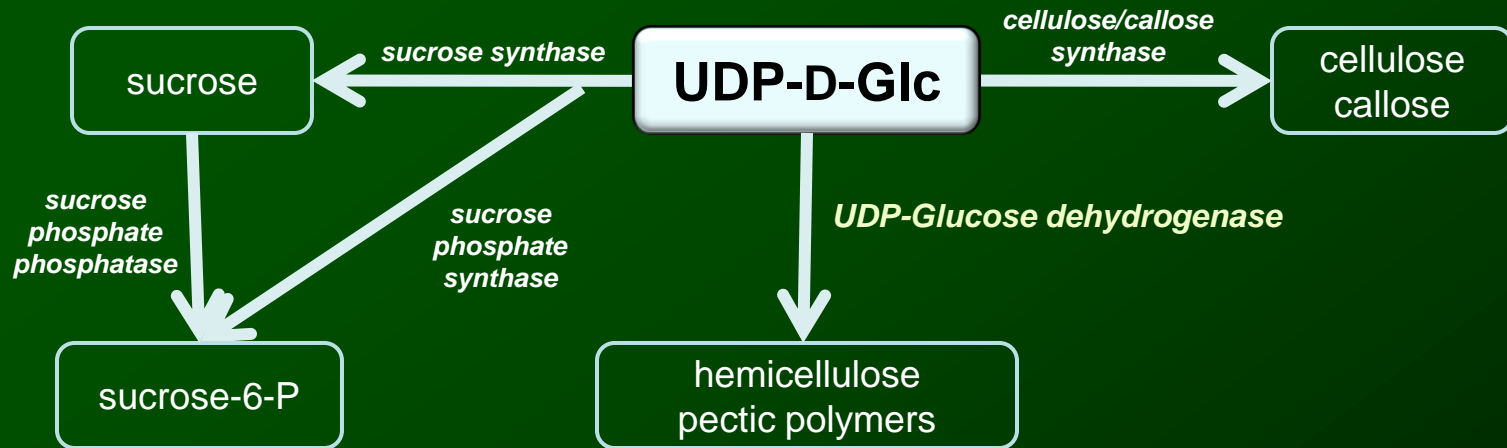
#### Storage Parenchyma Cell



● Enzymes/transporters targeted through transgenesis



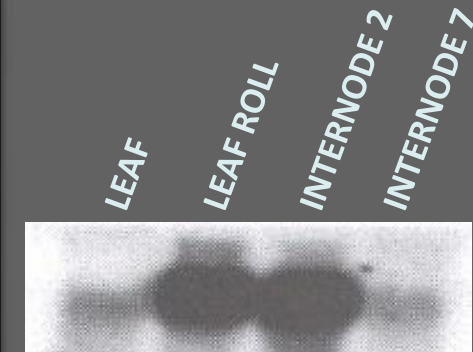
# UDP-GLUCOSE: A CENTRAL ROLE IN SUCROSE AND CELL WALL METABOLISM



ESTs Preferentially Expressed in Sugarcane Immature Internodal Tissue. ESTs were isolated by reciprocal **suppression subtractive hybridisations**.

Clone	Sequence homology/ match	Accession Number	Sequence Identity (%)	E Value
I2-6	<i>Glycine max</i> UDP-glucose dehydrogenase	U53418	78	9.7x10 <sup>-44</sup>
I2-7	<i>G. max</i> UDP-glucose dehydrogenase	U53418	92	4.2x10 <sup>-54</sup>
I2-13	<i>G. max</i> UDP-glucose dehydrogenase	U53418	96	1.4x10 <sup>-9</sup>
I2-155	<i>G. max</i> UDP-glucose dehydrogenase	T08818	90	2.0x10 <sup>-43</sup>
I2-160	<i>G. max</i> UDP-glucose dehydrogenase	Q96558	95	1.0x10 <sup>-39</sup>

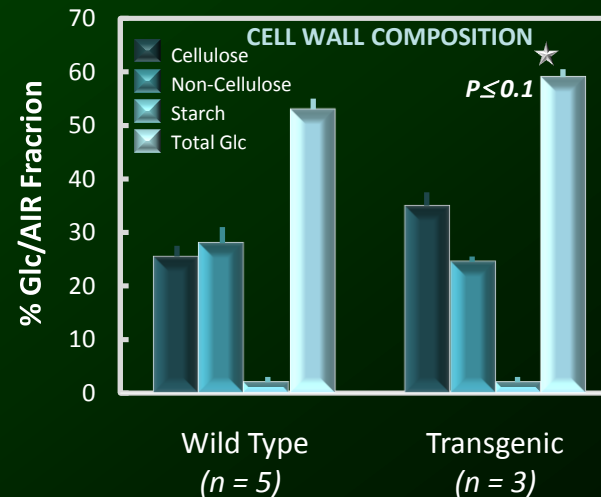
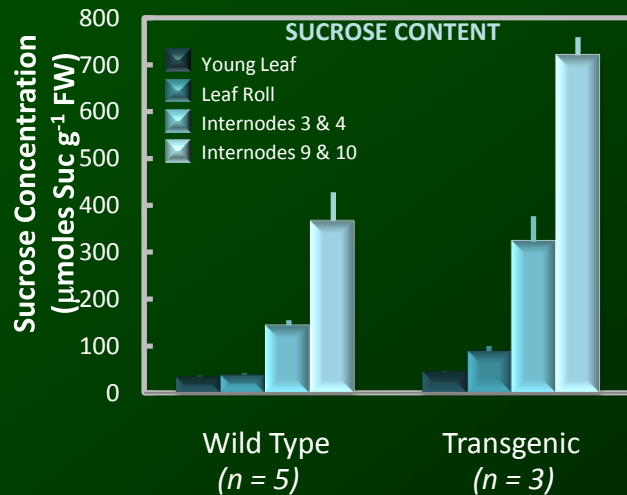
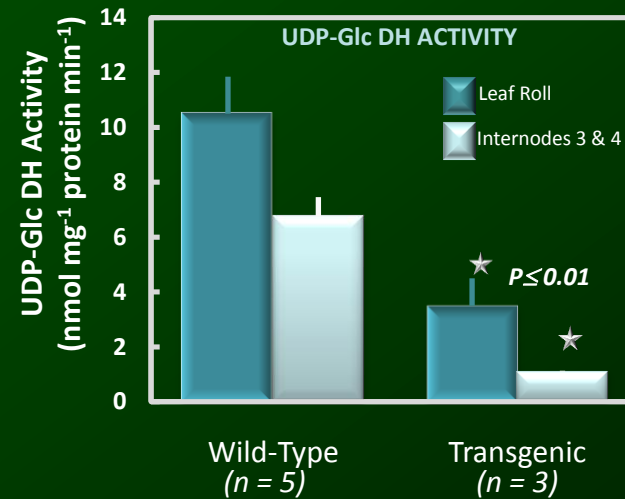
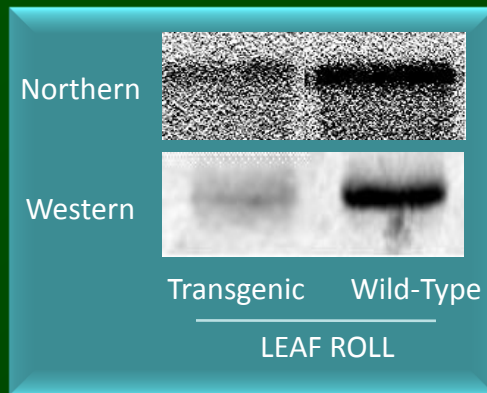
Expression of UDP-Glc DH  
Clone I2-160 in  
*Saccharum* spp hybrid  
NCo376



# EST Profiling & SSH: Provided Direction and Genetic Resources

## TARGETTING SUCROSE AND CELL WALL METABOLISM

### Down-Regulation of UDP-Glucose Dehydrogenase (Greenhouse Trials)



Internodes 9 + 10

# TARGETING SUCROSE AND CELL WALL METABOLISM

## Down-Regulation of UDP-Glucose Dehydrogenase (Greenhouse Trials)

Repression of UDP-Glc DH Activity

↑ [Uronic Acid]

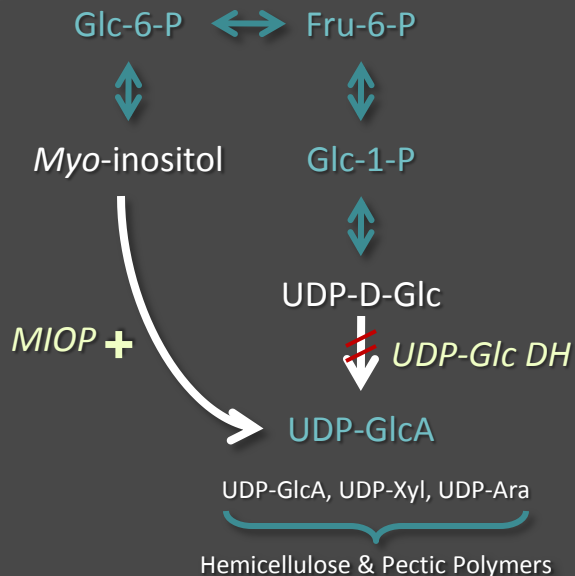
↑ [Total Glc] & Cellulose

↑ Arabinose : Galactose

↑ Pentose : Hexose (NS)

↑ Xylose : Galactose (NS)

Compensatory ↑ in Myo-Inositol Oxygenase Activity



Validation Under Field Conditions in Progress

### Lessons Learned:

- Confirmation of Plasticity of Primary Metabolism
- Necessity of Exploring Changes in Surrounding Metabolism

# SUGARCANE BIOLOGY: SOURCE-SINK RELATIONSHIPS

## 3 GENE SEQUENCES & THEIR EXPRESSION: UNDERSTANDING SYSTEMS

### Physiological processes that may participate in regulating sucrose accumulation

LEAF

Leaf photosynthetic rates & photosynthate partitioning/sequestration

TRANSPORT

Phloem loading & unloading (& leakage & recapture during transport)

STALK

Membrane transport (cell membrane, tonoplast)

Sucrose turnover (cycling, remobilization)

Allocation (glycolytic flux, fibre biosynthesis)

Sensitivity of source photosynthetic activity to sink demand

  
NEW FOCUS



## Evidence for supply-demand driven regulation of photosynthesis in sugarcane:

### End product repression of photosynthesis in sugarcane:

(Hartt & Burr, 1967; Waldron *et al.*, 1967 Bull & Tovey, 1974; Irvine, 1975; Amaya *et al.*, 1995; Allison *et al.*, 1997)

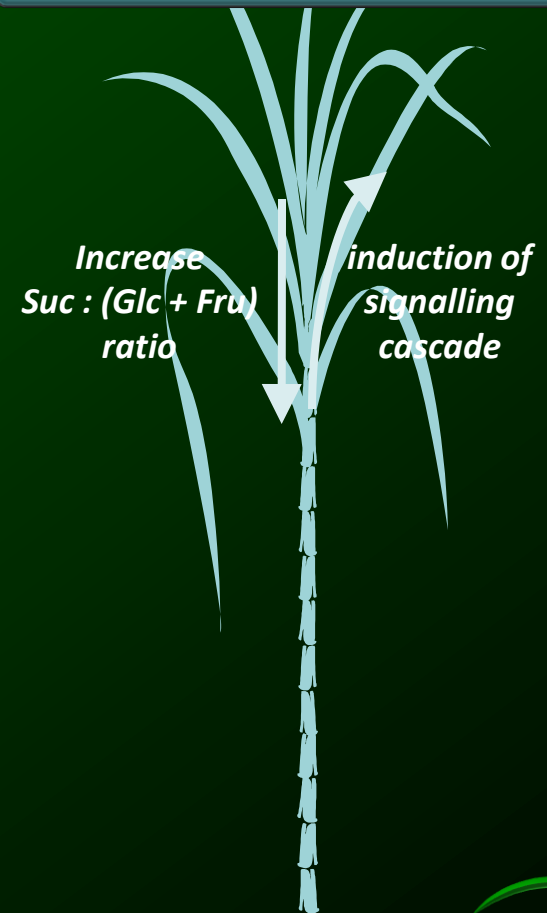
### Evidence of photosynthetic 'plasticity' in sugarcane:

(Wu & Birch, 2007)

### Well-documented evidence of source-sink feedback in other plants:

(Paul & Foyer, 2001; Paul & Pellny, 2003; Rolland *et al.*, 2006; Paul, 2007)

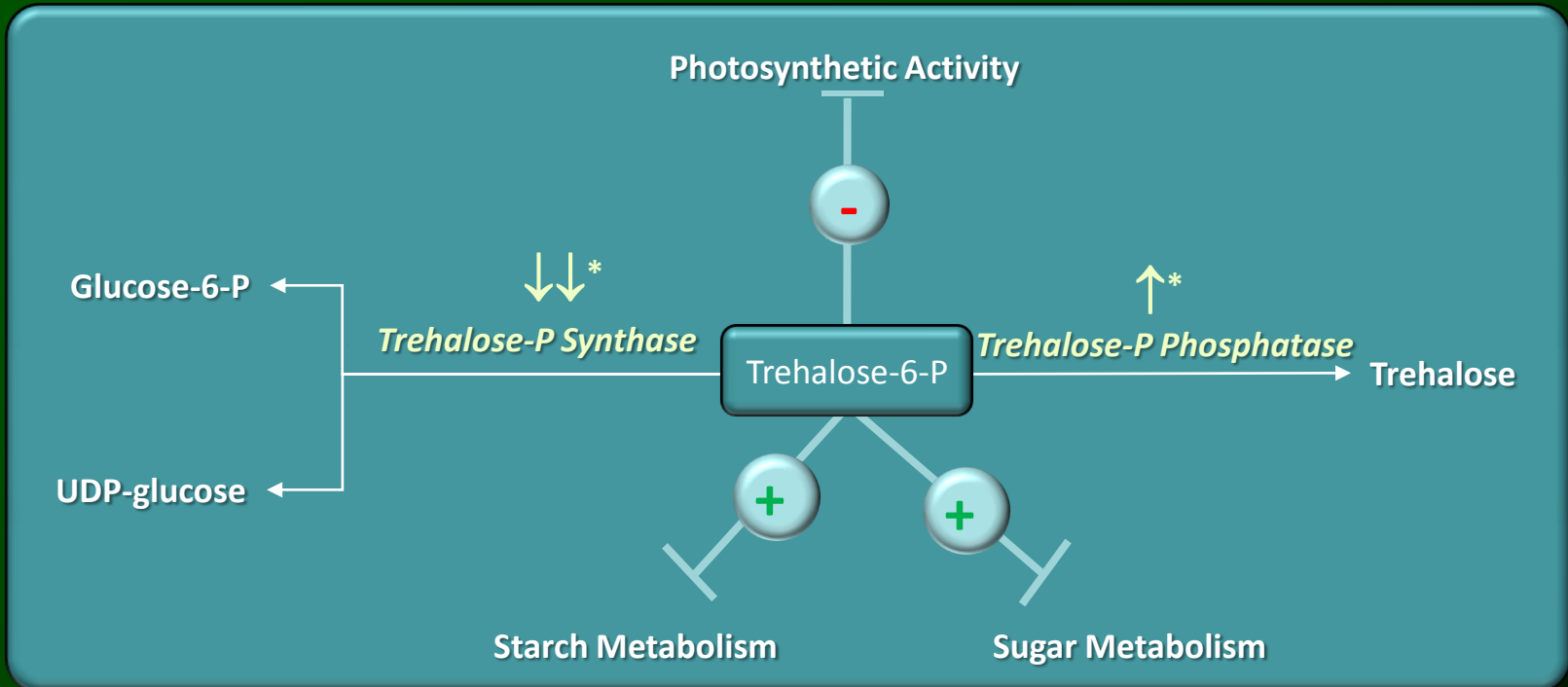
- Down-regulation of photosynthesis
- Induction of leaf physiological decline (senescence)



# Analysis of Potential Signalling Mechanism: Affymetrix Sugarcane Genome GeneChip

UP-REGULATED			DOWN-REGULATED		
Putative Gene Identity	Fold Change		Putative Gene Identity	Fold Change	
<b>Photosynthesis</b>					
			ATP synthase CF0 C Chain	0.20	↓↓
			ATP synthase CF1 beta chain	0.30	↓↓
			Cytochrome P540	0.10	↓↓↓
			Photosystem I subunit O	0.05	↓↓↓
			Phosphoglycerate kinase (chloroplastic)	0.30	↓↓
			RuBisCO large subunit-binding protein	0.40	↓
			RuBisCO small subunit	0.10	↓↓↓
<b>Sugar signalling, transport &amp; P<sub>i</sub> metabolism</b>					
Glucose-phosphate/phosphate transporter	9.7	↑↑↑	Trehalose-6-phosphate synthase	0.30	↓↓
Trehalose-phosphatase	3.6	↑			
Inorganic pyrophosphatase	4.4	↑↑			
Phosphate translocator	4.4	↑↑			
<b>Starch metabolism</b>					
1,4- $\alpha$ -glucan branching enzyme	2.0	↑			
ADP-glucose pyrophosphorylase	12.8	↑↑↑			
$\beta$ -amylase	5.2	↑↑			
Isoamylase	8.2	↑↑↑			
Phosphorylase	8.8	↑↑↑			

# Potential Central Role of Trehalose-6-Phosphate in Source-Sink Signalling



\* As per Affymetrix Sugarcane Genome GeneChip analysis

## PROSPECTS:

APPLICATIONS OF NEW TECHNOLOGIES & RESOURCES IN GENOMICS & FUNCTIONAL GENOMICS

**MARKER-ASSISTED  
BREEDING**

**SUGARCANE BIOLOGY**  
(Source-Sink Relationships)





# MARKER-ASSISTED BREEDING : CURRENT RESEARCH



## ICSB Projects

LD Map of SA breeding population

Identification of QTAs for yield component traits

Extended LD Map of SA population (DArT)

Extended reference genetic map of R570 population

Project 21

Project 26

### R570 REFERENCE MAP

MARKER	NUMBER
RFLP	211
SSR	140
AFLP	689
DArT	890
<b>TOTAL</b>	<b>1930</b>

### LD MAP

MARKER	NUMBER
AFLP	737
DArT	1906
Unique DArT	1263
Unique Markers	890

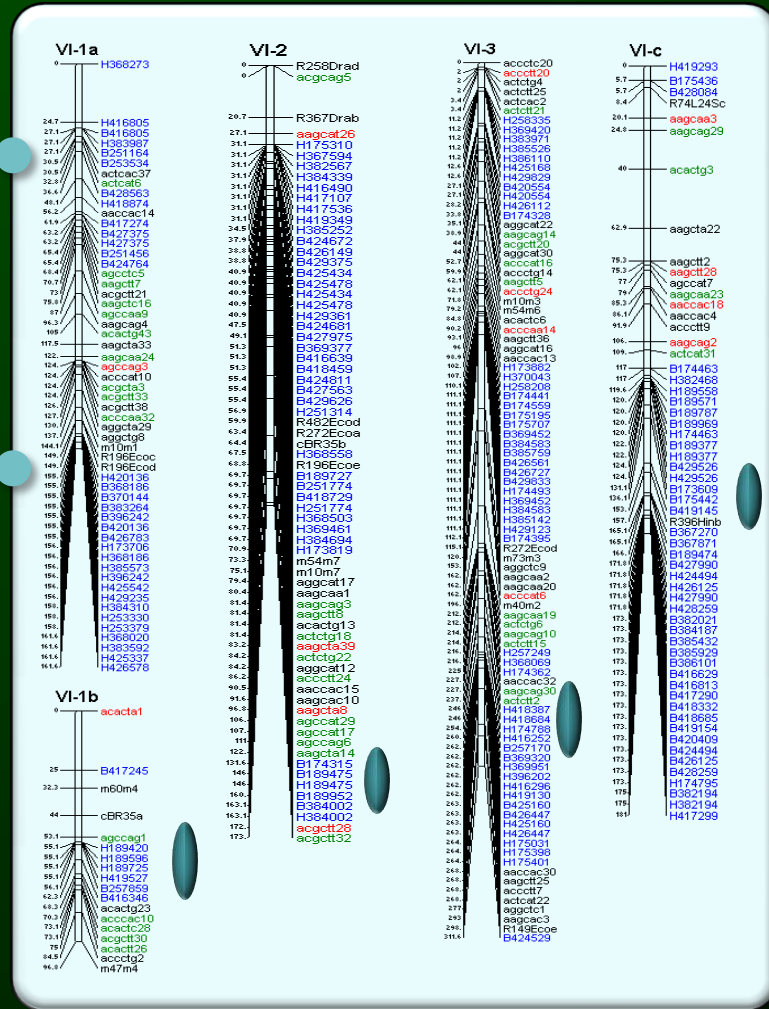
373 DArT markers common to both maps



# MARKER-ASSISTED BREEDING : CURRENT RESEARCH

Established position of markers from the LD map on the genetic map of R570

Identified potential QTLs for cane yield from LD Map





# MARKER-ASSISTED BREEDING : PROSPECTS

EXPLOIT

Existing sugarcane genetic maps (R570)

Established marker-trait associations (LD population)

Synteny amongst sugarcane & close *Saccharum* relatives

TO MINE

Available resources e.g. the genome sequences of *Sorghum bicolor* & *Brachypodium distachyon*

TO DEVELOP SEQUENCE-SPECIFIC MARKERS (e.g. SNPs) TO TARGET INDIVIDUAL ALLELES

Marker-Assisted Breeding

Transgenesis



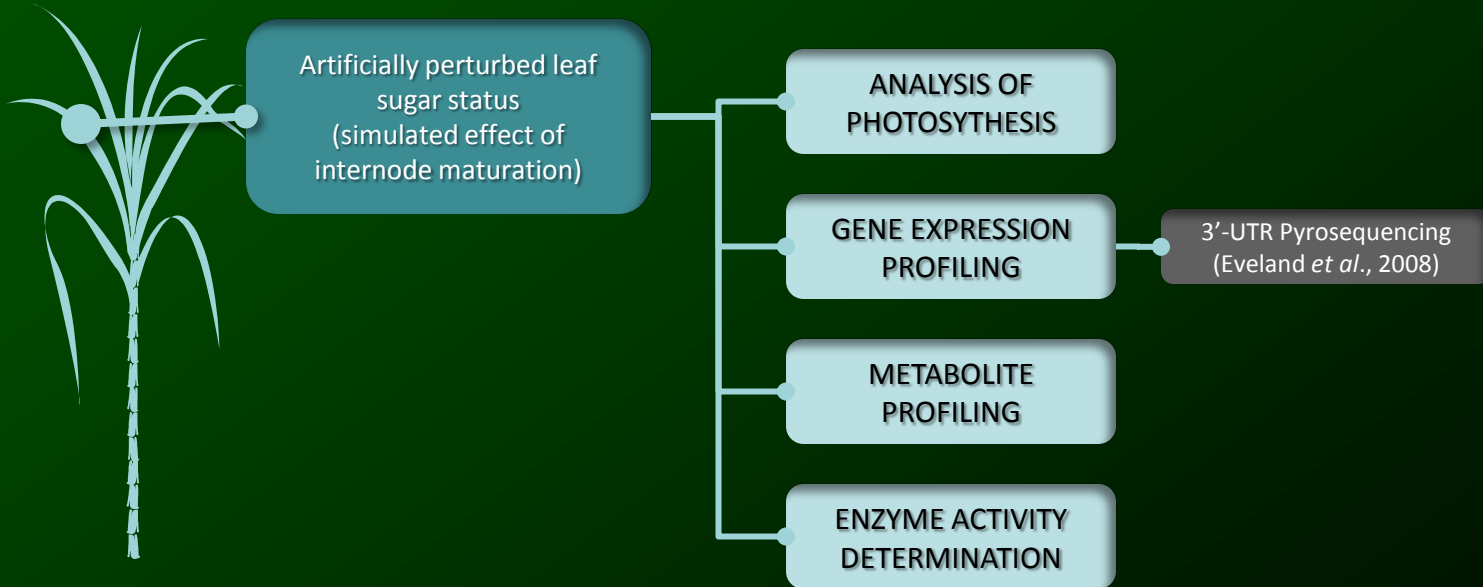


# SUGARCANE BIOLOGY (SOURCE-SINK RELATIONSHIPS) : CURRENT RESEARCH

## GOAL

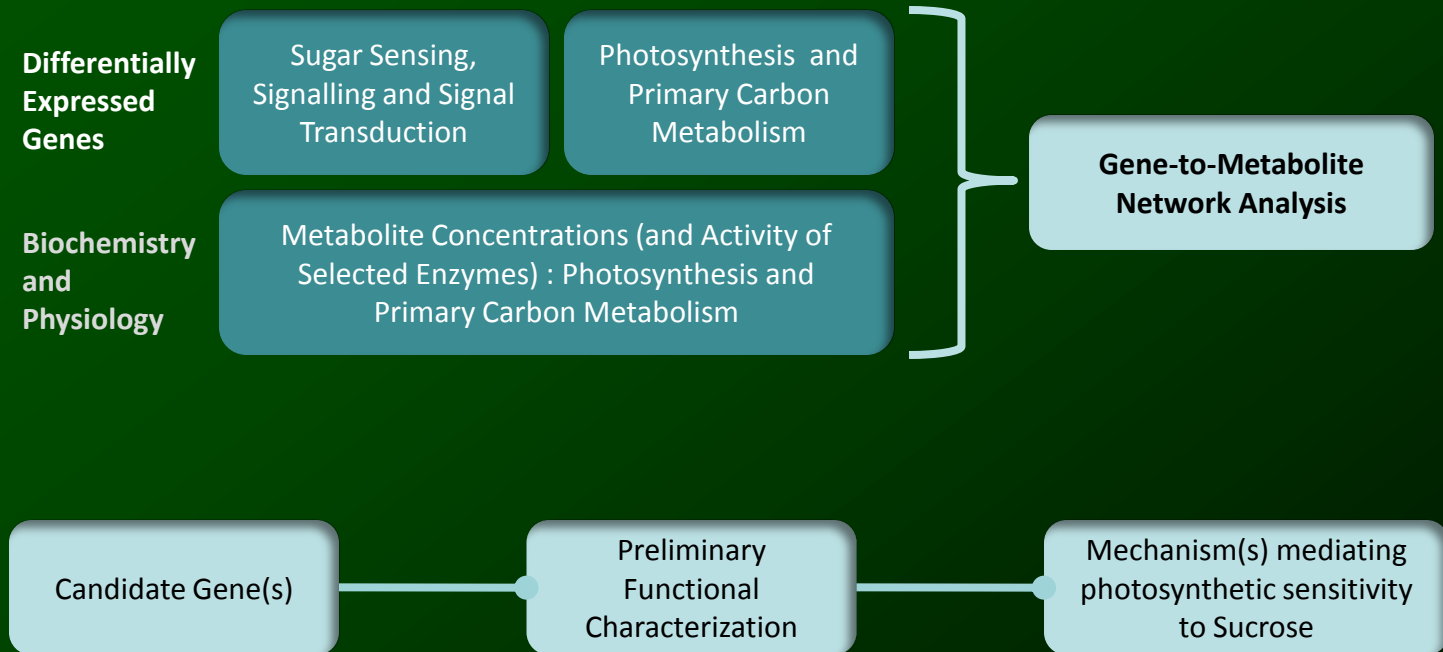
To deploy high-throughput technologies and bioinformatic resources to determine the molecular mechanism underlying the sensitivity of photosynthetic carbon metabolism and related metabolic pathways to fluctuating sugar concentrations in the leaves of sugarcane.

## APPROACH



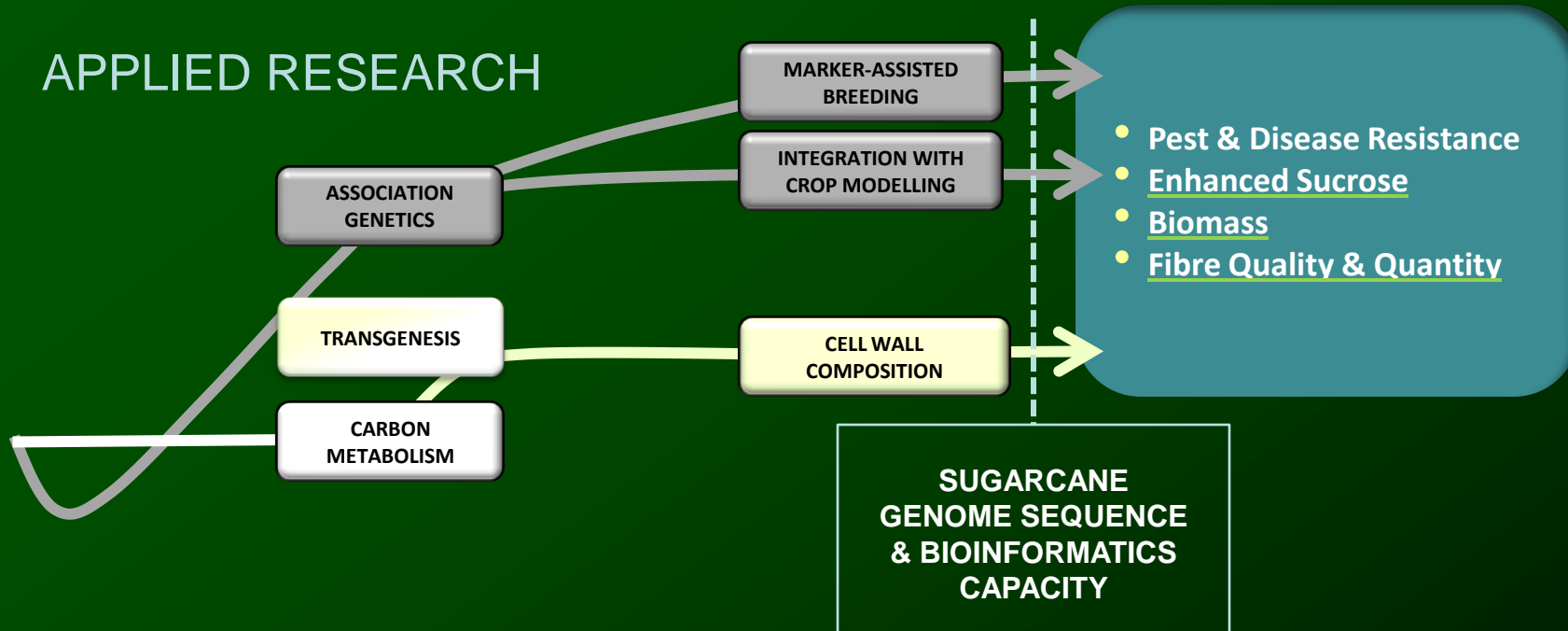


## Changes in the Leaf Associated with Internode Maturation (increased Suc : (Glc + Fru Ratio))

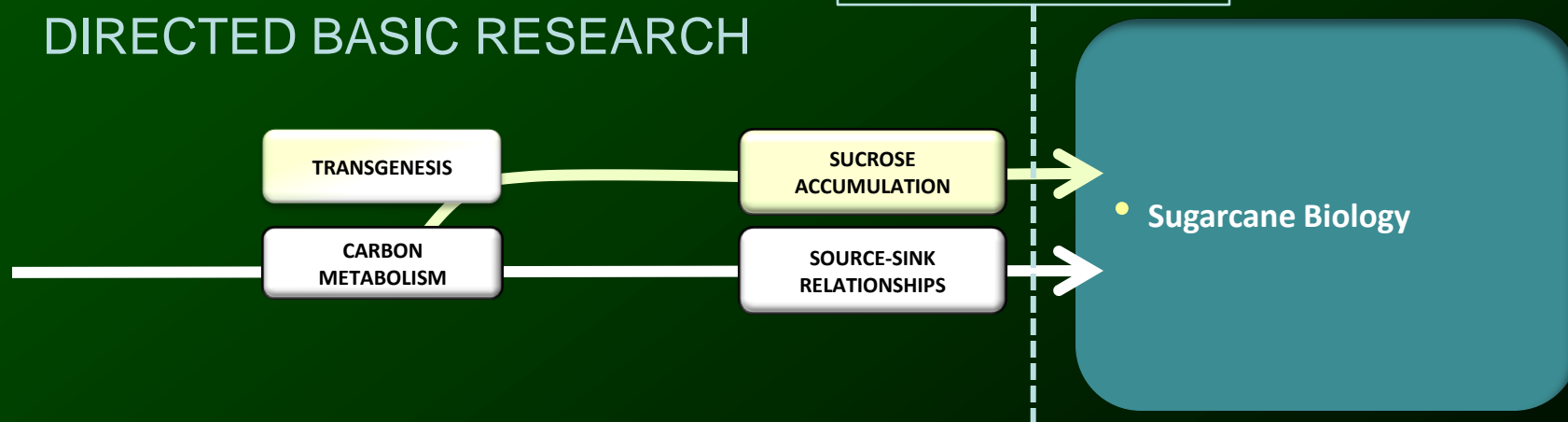


# CHALLENGES : GENOMICS & FUNCTIONAL GENOMICS

## APPLIED RESEARCH



## DIRECTED BASIC RESEARCH



Frikkie Botha  
Barbara Hockett  
Mike Butterfield  
Mike Cramer (University of Cape Town)  
Jens Kossmann (Institute of Plant Biotechnology)  
Alistair McCormick  
Deborah Sweby (formerly Carson)



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